

Giant Magnetoresistive Biosensor Array for Detecting Magnetorelaxation

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Abstract—In this paper, a time-domain magnetorelaxometry biosensing scheme is presented using giant magnetoresistive (GMR) sensors to measure the fast relaxation response of superparamagnetic magnetic nanoparticles (MNPs) in a pulsed magnetic field. The system consists of an 8×10 GMR sensor array, a Helmholtz coil, an electromagnet driver, and an integrator-based analog front-end needed to capture the fast relaxation dynamics of MNPs. A custom designed electromagnet driver and Helmholtz coil improve the switch-off speed to $>5 \text{ Oe}/\mu\text{s}$, limiting the dead zone time to $<10 \mu\text{s}$, and thus enables the system to monitor fast relaxation processes of 30 nm MNPs. A magnetic correlated double sampling technique is proposed to reduce sensor-to-sensor variation by 99.98% while also reducing temperature drift, circuit offset, and nonlinearity below the noise level. An optimum integration time is calculated and experimentally verified to maximize the SNR. Experiments with dried MNPs have shown successful relaxation detection, and immunoassay experiments have demonstrated their binding kinetics.

Index Terms—Electromagnet driver, magnetorelaxometry biosensing, magnetic correlated double sampling, magnetic nanoparticles, magnetoresistive sensors.

I. INTRODUCTION

RECENTLY, magnetic sensors that have been traditionally utilized in industrial applications are being used more and more in medical applications, such as point-of-care (POC) *in-vitro* diagnostics [1]–[15]. In magnetic biosensing, biomarkers (e.g., proteins, nucleic acids, cells, etc.) are tagged with magnetic nanoparticles (MNPs) and detected by magnetic sensors to diagnose and track the progression of diseases [4]–[15]. Fig. 1(a) shows one such example where a magnetic immunoassay is utilized for protein detection. The biomarker of interest is selectively captured by an antibody immobilized on the surface of the sensor and subsequently bound to a detection antibody conjugated with a MNP. By quantifying the number of MNPs tethered to the magnetic sensor, one can infer the concentration of biomarkers present in the sample.

Several types of magnetic sensors have been successfully used to detect magnetically labelled biomarkers. Nuclear mag-

Manuscript received August 30, 2016; revised December 26, 2016; accepted February 9, 2017. Date of publication June 2, 2017; date of current version July 26, 2017. This work was supported in part by the National Science Foundation under Grant ECCS-1454608 and in part by Qualcomm. This paper was recommended by Associate Editor M. Zaghoul.

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Digital Object Identifier 10.1109/TBCAS.2017.2682080

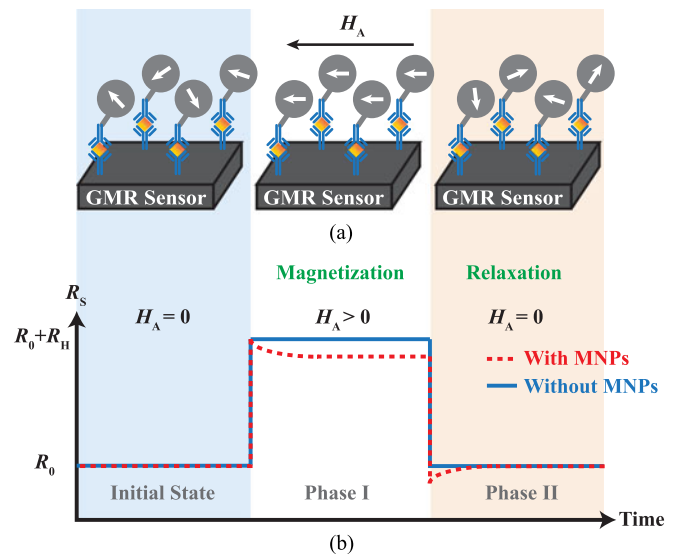


Fig. 1. Magnetorelaxometry biosensing concept. (a) A sandwich complex of antibody, antigen, antibody, and MNP bound on the sensor surface. (b) GMR sensor response due to a pulsed magnetic field.

netic resonance (NMR) sensors [3], [4] measure the bulk spin-spin relaxation time (T_2) of water molecules to indirectly detect magnetic clusters. Hall-effect sensors [9] measure the Hall voltage arising from bound magnetic beads. MNPs have also been shown to induce frequency shifts in LC resonators [10]. Among the many magnetic sensors available, giant magnetoresistive (GMR) devices stand out for this application due to their high transduction efficiency, particularly at low magnetic fields [16], [17]. Traditionally, the MNPs tethered to the sensor surface have been detected by the local perturbation in the magnetic field [18]–[21]. While very sensitive, this technique requires high magnetic field uniformity, complex modulation schemes to deal with the low signal to baseline ratio, and long measurement time needed to reduce the noise.

Here, we propose an alternative temporal detection scheme, magnetorelaxometry (MRX), using a pulsed magnetic field, as shown in Fig. 1. Without an applied magnetic field, the magnetic moments of the superparamagnetic MNPs are randomly distributed resulting in zero net magnetic field. In Phase I, an external magnetic field H_A is applied, magnetizing and aligning the MNPs with the applied field. The underlying GMR sensor detects both the external field and the stray field from the MNPs, as denoted by the change in resistance. In Phase II, H_A is quickly removed resulting in Néel and/or Brownian relaxation

of the MNPs. For Brownian relaxation, the magnetization vectors undergo rigid rotation of the whole particle due to Brownian motion [22]. However, the MNPs here are tethered to the surface and thus have limited movement [23]. Instead, the MNPs must undergo Néel relaxation, where internal domain movement results in gradual randomization of the moment [24]. Note that the GMR sensor also exhibits Néel relaxation, but is orders of magnitude faster than the MNPs, allowing one to use the relaxation signature to detect the number of tethered MNPs. Compared with traditional magnetometry, which measures the absolute sensor resistance difference during Phase I, MRX shifts the burden from accuracy in amplitude to accuracy in time, relaxing the requirement on external field uniformity. Furthermore, since the relaxation of the MNPs is usually fast (nanosecond to millisecond time scale), MRX has both a shorter measurement time and the flexibility to tradeoff sensitivity with measurement time by adjusting the number of averaging cycles. Lastly, MRX is capable of distinguishing different types of MNPs on multi-labelled biomarkers due to their different relaxation times.

The MNP magnetization as a function of time can be written as follows [25]:

$$M_N(t) = C \cdot \ln \left(1 + \frac{t_C}{t} \right) \quad (1)$$

where t is the time after switching off the applied field, t_C is the characteristic time that depends on the magnetization time and the field strength, and C is a scaling factor that depends on the surface coverage, magnetic viscosity, and initial magnetization.

This relaxation phenomenon can be measured either in the frequency- or time-domain. Frequency-domain MRX measures the complex magnetic susceptibility as function of frequency in an alternating magnetic field [26], whereas time-domain MRX directly measures the relaxation signal in response to a pulsed magnetic field [27], [28]. One of the significant challenges in time-domain relaxation is the need to collapse the applied magnetic field faster than the relaxation of the MNPs, which can be in the nanosecond to millisecond range depending on the size and material composition of the MNPs [29]–[31].

In this paper, a time-domain integrator-based MRX biosensing system is proposed. A magnetic correlated double sampling (MCDS) technique is applied to eliminate sensor-to-sensor variation, temperature drift, circuit offset, and non-linearity. We also designed an ultrafast electromagnet driver, which uses a large negative voltage to discharge the electromagnet and improve the switch-off speed to $>5 \text{ Oe}/\mu\text{s}$.

The rest of this paper is organized as follows: Section II describes the proposed circuit architecture, Section III provides implementation details, and Section IV discusses testing methods and measurement results. Lastly, conclusions are drawn in Section V.

II. SYSTEM ARCHITECTURE AND CIRCUIT DESIGN

A. System Architecture

A block diagram of the proposed MRX detector is shown in Fig. 2. A PC provides a user-defined magnetization time and magnetic field amplitude to the PC-FPGA interface. The

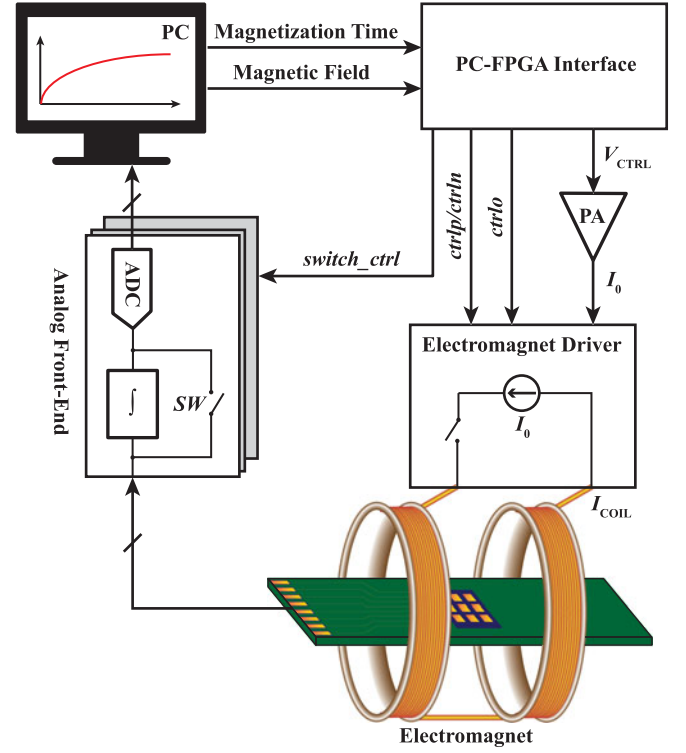


Fig. 2. Block diagram of the proposed relaxation detection system.

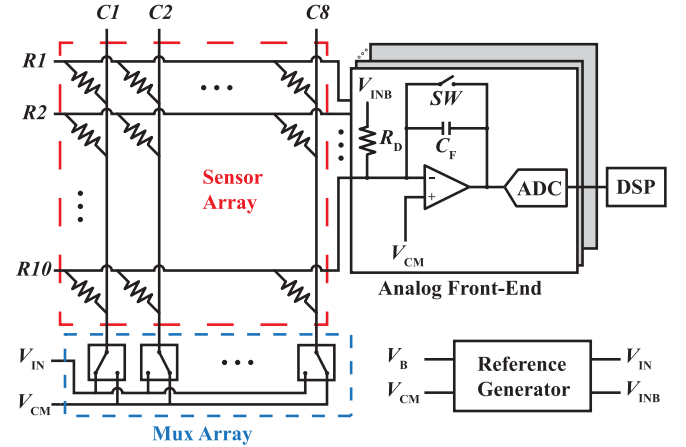


Fig. 3. Sensor array and readout architecture.

FPGA controls the timing of the electromagnet and readout circuitry. The power amplifier (PA) provides a DC current I_0 to the electromagnet driver that delivers a pulsed current I_{COIL} to the Helmholtz coil, generating a pulsed magnetic field for the sensor chip located in the center. The analog front-end (AFE) captures the resulting sensor response and transfers the data to the PC for post-processing.

The architecture of the AFE is illustrated in Fig. 3. The 8×10 GMR sensor array is arranged in a matrix with shared columns and rows. All the columns are connected to a column switching matrix that connects one column to V_{IN} and all others to a common-mode voltage, V_{CM} . Each of the rows is connected to a

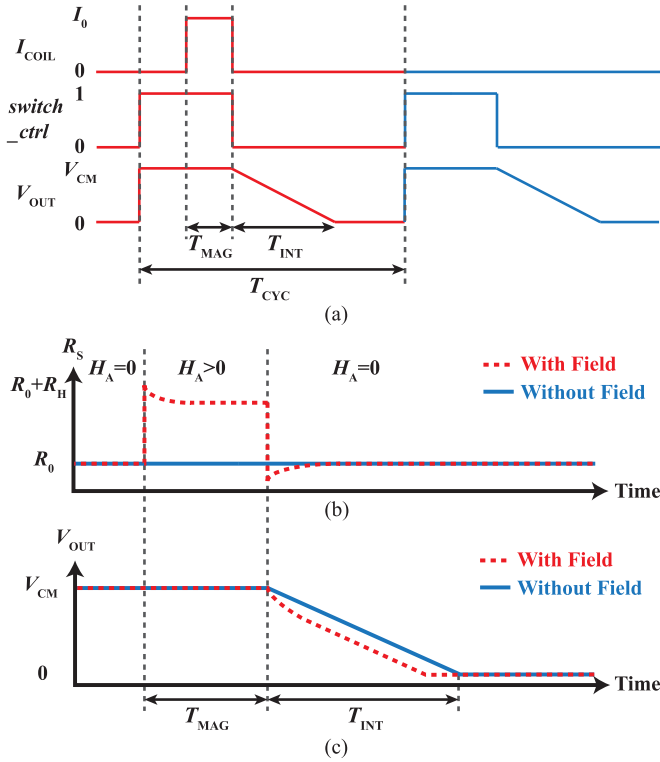


Fig. 4. (a) Timing diagram of coil current, integrator switch control, and integrator output voltage. (b) Sensor response and (c) corresponding integrator output using the MCDS scheme with and without a field applied.

readout circuit consisting of a switched integrator, an antialiasing filter, and an analog to digital converter (ADC). The reference generator produces voltages V_{IN} and V_{INB} as $V_{CM} + V_B$ and $V_{CM} - V_B$, which are used in the switched integrators as will be discussed later.

B. Magnetic Correlated Double Sampling

Detecting the minute relaxation signal in the presence of a large background signal is challenging. Traditional techniques, such as a Wheatstone bridge [17]–[19], require matched reference sensors to reject the background signal as common-mode. However, this approach is very sensitive to sensor-to-sensor variation and temperature drift, both of which are challenging to control for GMR sensors [32]. Moreover, circuit imperfections, such as amplifier offset and non-linearity, can further limit the sensitivity of the whole system. To extract the relaxation signal from these error sources, a MCDS technique is applied, as shown in Fig. 4. The magnetic field is initially off and the integrator reset switch SW is closed. Then, the magnetic field is turned on for a duration of T_{MAG} , while SW remains closed (leaving the op-amp in unity gain configuration), therefore $V_{OUT} = V_{CM}$. Next, the magnetic field is turned off and the reset switch is opened simultaneously, resulting in V_{OUT} decaying. The integration time T_{INT} is determined by the sensor resistance R_0 and thus can be different due to sensor-to-sensor variation. This process is repeated in the subsequent cycle, except that the magnetic field remains off. Thus, the first cycle has the magnetic relaxation signal whereas the second does not.

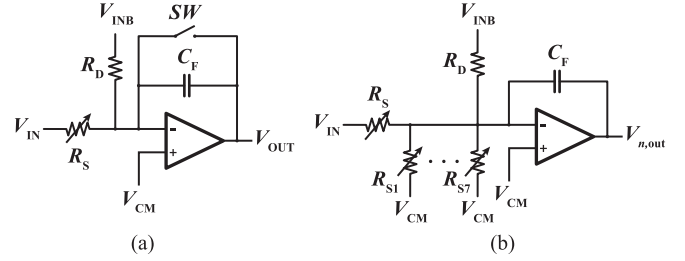


Fig. 5. (a) Switched integrator schematic and (b) equivalent schematic for noise analysis.

The sensor response and V_{OUT} in both cycles are shown in Fig. 4(b) and (c), respectively. When the magnetic field is turned off, the sensor resistance returns to the nominal resistance R_0 due to the relaxation of the MNPs. Therefore, instead of linear decay of V_{OUT} in the case without the field, V_{OUT} is distorted at the beginning of integration and returns to linear decay when the relaxation of the MNPs is completed. By subtracting V_{OUT} with and without the field, the integrated relaxation signal can be extracted.

This technique removes the sensor-to-sensor variation since each sensor is compared to itself instead of a reference sensor. While temperature may cause sensor resistance drift over time, the change is very slow compared to the period of cycle T_{CYC} . Therefore, MCDS is able to reduce the effect of temperature drift dramatically without the need for other temperature compensation techniques [32]. By selecting T_{CYC} as a multiple of the power line frequency, the 60 Hz interference in the two adjacent cycles will be in phase and thus rejected as well. This technique is also effective at eliminating errors such as the circuit offset and non-linearity since these errors are consistent in every cycle.

C. Switched Integrator

In prior work, time-domain relaxation measurement techniques used amplifiers to detect the relaxation signal with Hall effect sensors [27], [28]. However, since the relaxation signal is largest at the time when the external field is turned off and gradually decreases with time, traditional amplifier-based relaxometry requires a high sampling-rate ADC to capture the dynamics. Since a high-resolution ADC is often needed to reduce the quantization error, integrators are applied to relax the bandwidth requirement of the ADC by integrating the relaxation signal with time, thus the integrated signal increases with time. The switched integrator circuit is shown in Fig. 5(a). It consists of an op-amp with capacitive feedback, a reset switch, and a current cancellation resistor at the input. Since GMR sensors typically have magnetoresistance ratios between 6–25% [17], there will be a substantial portion of the current flowing into the integrator that does not respond to the magnetic field and would limit the dynamic range. To remedy this issue, we have added a cancellation network that consists of resistor R_D that bleeds off part of the baseline current allowing us to use a smaller feedback capacitor C_F for the same integration time T_{INT} . R_D

is selected $\sim 10\%$ lower than the nominal sensor resistance R_0 to account for process variation, and $V_{IN} = V_{CM} + V_B$ and $V_{INB} = V_{CM} - V_B$ where V_{CM} is a fixed common-mode voltage and V_B is an adjustable bias voltage. Consequently, in the integration phase, the output voltage V_{OUT} can be written as follows:

$$V_{OUT} = V_{CM} - \frac{1}{C_F} \int \frac{V_B}{R_0 + \Delta R(t)} - \frac{V_B}{R_D} dt \quad (2)$$

where t is the time, and $\Delta R(t)$ represents the sensor response due to the MNP relaxation.

The relaxation signal after MCDS, ΔV_{OUT} , can be derived as follows based on the assumption that $R_0 \gg \Delta R(t)$:

$$\begin{aligned} \Delta V_{OUT} &= V_{OUT}|_{\text{without field}} - V_{OUT}|_{\text{with field}} \\ &= \frac{V_B}{C_F} \int \frac{1}{R_0 + \Delta R(t)} - \frac{1}{R_0} dt \\ &\cong -\frac{V_B}{R_0^2 C_F} \int \Delta R(t) dt. \end{aligned} \quad (3)$$

The equivalent schematic for noise analysis is shown in Fig. 5(b). Since the parasitic capacitance at the inverting node of the op-amp is much smaller than C_F , reset noise of the integrator is negligible [33]. In addition to the sensor under test R_S , all other sensors $R_{S1} - R_{S7}$ in the same row contribute to the noise due to the matrix configuration. The noises contributed by V_{CM} , V_{IN} , and V_{INB} are experimentally verified to be negligible due to their low bandwidth. Hence, the output-referred noise can be derived as follows:

$$\begin{aligned} v_{n,out}^2 &= \left(\frac{1}{2\pi f C_F} \right)^2 \left(8i_{n,Rs}^2 + \frac{4kT}{R_D} \right) \\ &+ \left(\frac{9}{2\pi f R_S C_F} + 1 \right)^2 v_{n,op}^2 \end{aligned} \quad (4)$$

where $i_{n,Rs}$ represents the current noise contributed by each sensor, $v_{n,op}$ represents the voltage noise contributed by the op-amp, k is the Boltzmann constant, T is the temperature, and f is the frequency. Both $i_{n,Rs}$ and $v_{n,op}$ include white and $1/f$ noise.

D. Electromagnet Driver

The finite switching time of the magnetic field results in a deadzone where measurements cannot be taken. To minimize the deadzone time to detect MNPs with fast relaxation times, the switch-off response of magnetic field needs to be as fast as possible. However, this requirement is at odds with typical methods of generating magnetic fields that result in large inductances. Both stripelines [34] and miniaturized coils [35], [36] have been used for achieving a fast switch-off response due to their low inductance. However, stripelines are incapable of generating large magnetic fields (3.2 Oe in [34]) and lose control over the field uniformity, and miniaturized coils require complex micromachining techniques. Helmholtz coils, on the other hand, generate homogeneous in-plane fields, are easy to fabricate, and facilitate proper sensor alignment due to their geometry [37]–

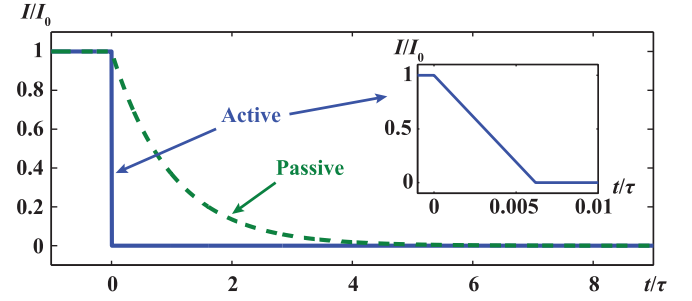


Fig. 6. Comparison of energy dissipation techniques.

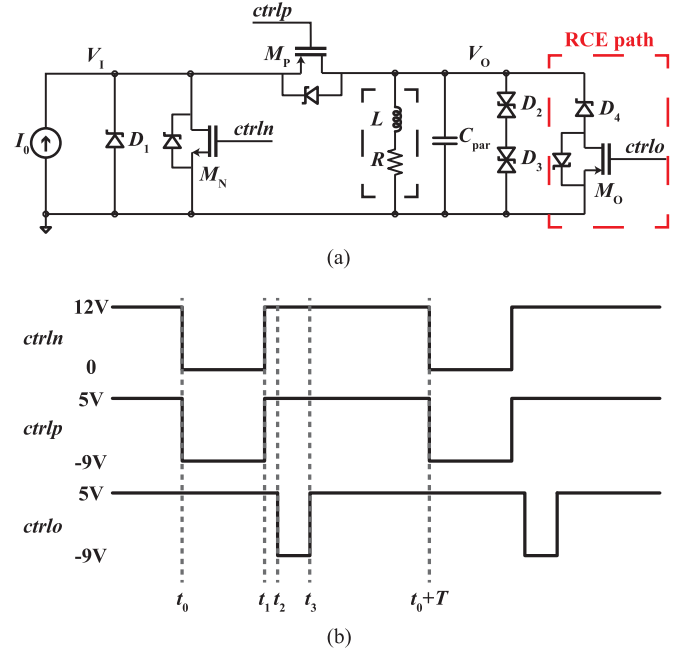


Fig. 7. (a) Schematic of electromagnet driver implementing active discharge. (b) Timing diagram of control signals.

[39]. However, the large inductance of a Helmholtz coil limits the bandwidth, making the fast switching difficult.

There are two methods to discharge the stored energy in an electromagnet: passively and actively. To passively discharge the electromagnet, both sides of the electromagnet are placed at the same potential and the current in the electromagnet can be written as: $I(t) = I_0 \exp(-t/\tau)$, where I_0 is the initial current, $\tau = L/R$ is the time constant, and t is time. Given that Helmholtz coils usually have large L and small R by design, this results in a large τ and slow decay of the current. Alternatively, the energy can be actively removed by connecting a negative voltage source V , accordingly $I(t) = I_0 - (V/L)t$. With a large enough V , this technique can rapidly remove the energy stored in the electromagnet within 0.006τ , $\sim 1,000\times$ faster (Fig. 6).

Based on the active discharge technique, a novel electromagnet driver was designed, as shown in Fig. 7(a). To the first order, the electromagnet can be modeled as an inductor in series with a resistor. The driver is operated in two phases and controlled by pulse width modulation (PWM) signals $ctrlrn$ and $ctrlrp$ through power MOSFET switches M_N and M_P . The reverse current

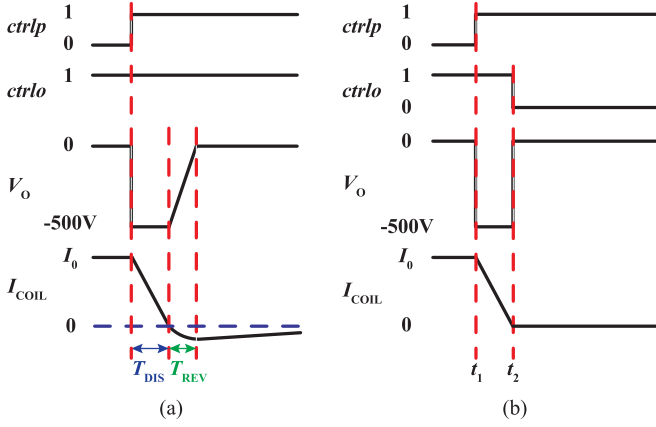


Fig. 8. (a) Reverse current due to parasitic capacitance. (b) RCE path eliminates the reverse current.

elimination (RCE) path, which consists of M_O , $ctrlp$, and D_4 , will be discussed later.

As shown in Fig. 7(b), during the charge phase ($t_0 - t_1$), $ctrlp$ and $ctrlp$ are low, therefore M_N is off and M_P is on. A DC current I_0 flows into the electromagnet through M_P . To discharge the electromagnet (at time $t_1 - t_0 + T$), $ctrlp$ and $ctrlp$ are high, therefore M_N is on and M_P is off. Transient voltage suppression (TVS) diodes D_2 and D_3 clamp V_O at a negative voltage of -500 V, which generates a low impedance path to rapidly remove the stored energy. While the negative voltage should be as large as possible to achieve a fast switch-off response, it is limited by both the electromagnet breakdown voltage and the drain-source breakdown voltage of M_P . Meanwhile, I_0 flows to ground through M_N , and diode D_1 protects M_N and the current source from large transient voltage spikes.

The proposed topology can speed up the switch-off response significantly, but comes with a reverse current, as shown in Fig. 8(a). After the energy stored in the coil has been fully dissipated, V_O is expected to rise back to 0 V. However, the parasitic capacitance C_{par} from the electromagnet, TVS diodes, MOSFET M_P , and PCB prevents V_O from instantly returning to 0 V. Since there is no external source to charge C_{par} , current is pulled out of the coil during T_{REV} . Therefore, after I_{COIL} drops down to 0 A, the negative V_O continues “discharging” the electromagnet, resulting in a reverse current whose magnitude keeps increasing until V_O reaches 0 V. Making this issue worse, as $|V_O|$ decreases from 500 V to 0 V, the low-impedance path generated by D_2 and D_3 is broken. The reverse current must be dissipated as heat by the electromagnet resistance and thus lingers for a comparatively long time. This phenomenon was observed experimentally and was also discussed in [40]. In our application, this reverse current and the associated magnetic field is not acceptable as it magnetizes the MNPs, instead of allowing them to relax to equilibrium.

Our solution is to reduce T_{REV} by forcing V_O to 0 V. To facilitate this, an RCE path is added at the output node in parallel with the electromagnet and TVS diodes, as shown in Fig. 7(a). The RCE path operates as follows: as soon as the energy in the electromagnet is fully dissipated, M_O is turned on, pro-

viding an alternative low-impedance path to charge C_{par} . This pulls V_O back to 0 V quickly, eliminating the reverse current. D_4 is needed to prevent unwanted current flow during the charge phase. The signal timing is illustrated in Fig. 8(b). It should be noted that the turn-on time (at $t_1 - t_2$) is nearly equal to the coil switch-off time, given that the rise and fall time of these signals are similar as confirmed through simulation and verified experimentally.

The timing of $ctrlp$ is critical and requires a sub-microsecond resolution for microsecond field switching times. Early turn-on causes V_O to start returning to 0 V too early when the energy stored in the electromagnet has not been fully dissipated. Without a large negative V_O , the residual current will decay exponentially with time constant of L/R , resulting in a long switch-off time. On the other hand, if $ctrlp$ arrives late, the reverse current cannot be fully removed.

While inductor-current zero-crossing detection methods have been demonstrated to achieve high-accuracy timing control [41], [42], they all require additional detection circuits. In our case, the zero-crossing detector is not needed since the correct timing can be derived as follows:

$$t = t_{os} + \frac{I_0 L}{|V_m|} \quad (5)$$

where t_{os} is the offset time, which is determined by the circuit delay, I_0 is the current flowing into the electromagnet, L is the electromagnet inductance, and V_m is the output voltage during switching-off, which is 500 V in our design.

For a given electromagnet, L and V_m are fixed. To use (5) to calculate the turn-on delay t based on a given current I_0 , the offset time t_{os} , which is highly dependent on the electromagnet parasitic capacitance, must be measured. The measurement procedure is as follows:

- 1) Choose a current value I_0 , turn M_O off, and measure how long it takes I_{COIL} to drop to 0 A.
- 2) With this time as M_O turn-on time, verify that the reverse current has been eliminated.
- 3) Calculate t_{os} using the known parameters.

This calibration procedure only needs to be done once per electromagnet to determine t_{os} , after which the timing can be calculated for different current values.

In addition to the electromagnet driver, optimizing the Helmholtz coil is critical to further improve the switch-off speed. The magnetic field generated by a Helmholtz coil and the inductance of the Helmholtz coil can be derived as follows:

$$H = 10^4 (0.8)^{2/3} \frac{\mu_0 n I}{R} \quad (6)$$

$$L = \frac{2R^2 n^2}{9R + 10W} \quad (7)$$

where H is the magnetic field intensity in Oe, L is the inductance, R is the radius of each coil, n is the number of turns, μ_0 is the permeability of free space, and W is the distance between the two coils. By combining (5)–(7), the switch-off time can be derived as follows:

$$t = t_{os} + \frac{2R^3 H n}{10^4 (0.8)^{2/3} \mu_0 V_m (9R + 10W)} \quad (8)$$

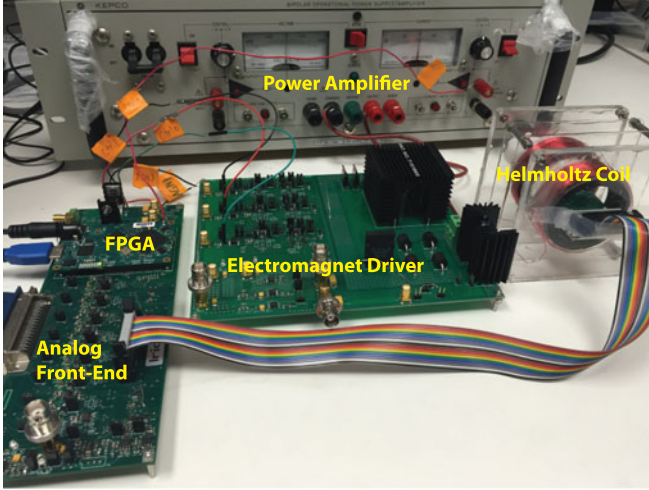


Fig. 9. Photograph of the measurement setup.

Therefore, for a fixed magnetic field, fewer turns, and/or a smaller radius result in faster switching time. However, the minimum radius is dictated by the sensor size. Fewer turns results in lower inductance and larger current with the maximum current usually limited by the heat dissipation.

III. SYSTEM IMPLEMENTATION

This system was implemented using an FPGA (Opal Kelly XEM6310), a National Instruments Data Acquisition Card (DAQ, NI PCIe-6351), a power amplifier (Kepco BOP 36-12ML), a custom designed Helmholtz coil, and two printed circuit boards (PCBs), as shown in Fig. 9. The DAQ has 8 analog input channels with 16-bit ADCs and 2 analog output channels with 16-bit DACs. The input channels were used to acquire the outputs of the integrators and the output channels were used to generate V_B for the integrator bias voltage and V_{CTRL} for the PA control voltage. The PA was set to constant current mode and can provide a DC current of up to 12 A. The FPGA was controlled through USB by MATLAB.

The electromagnet driver and the AFE were implemented on separate PCBs to isolate ground currents. The electromagnet driver has low voltage control circuitry on the left side and high power driving circuitry on the right side, separated by a thick and wide ground layer. The critical P-Channel MOSFET M_P on the electromagnet driver is a PolarP Power MOSFET (IXYS IXTK32P60P), which can handle 32 A and has a breakdown voltage of 600 V.

The sensor chip (MagArray, Inc.) has an 8×10 array of GMR sensors, each of which is $125 \mu\text{m} \times 125 \mu\text{m}$. It was positioned in the center of the electromagnet and connected to the AFE with a ribbon cable. The op-amps (Analog Devices, Inc. AD8655) for the switched integrators have rail-to-rail input/output. The feedback capacitors C_F are $0.68 \mu\text{F}$ film capacitors, since film capacitors have a lower voltage coefficient than ceramic capacitors. The discharge resistors R_D were set to $1.8 \text{ k}\Omega$ slightly larger than the sensor range.

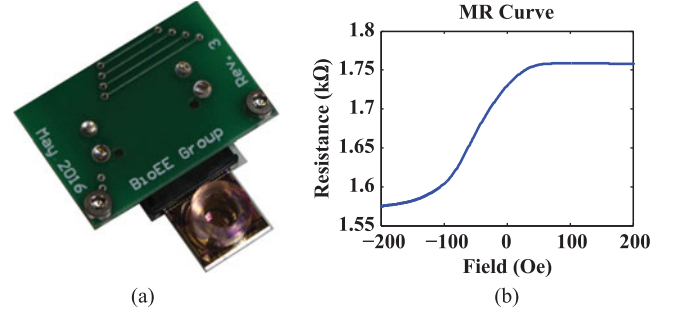


Fig. 10. (a) Photograph of the GMR sensor chip. (b) Measured magnetoresistance curve of the GMR sensor.

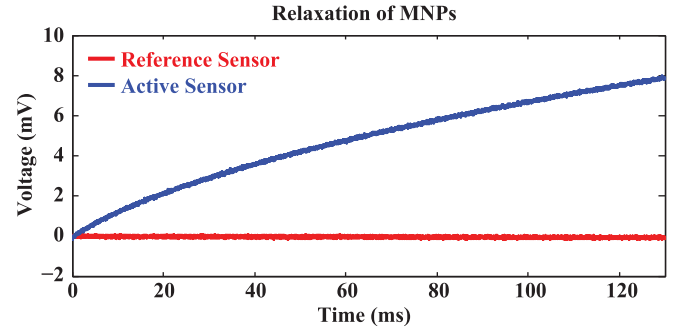


Fig. 11. Relaxation signal of dried MNPs.

IV. EXPERIMENTAL RESULTS AND DISCUSSION

The system was thoroughly characterized, first by verifying the sensors and measuring the noise, which allows further optimization for maximum SNR. We then characterized the ultrafast switching and RCE technique using two different electromagnets. Moreover, it was verified that the MCDS technique improved the system performance by reducing non-linearity and temperature drift. Lastly, the relaxation signal of commercially available MNPs was measured and an immunoassay was run.

A. GMR Sensor Characterization

The GMR sensors were characterized without MNPs by measuring the magnetoresistance (MR) curve. The external field was swept from -200 Oe to 200 Oe at a rate of 1 Oe/s . The sensor resistance as a function of field is shown in Fig. 10. The nominal resistance in the array ranged from $1.6 \text{ k}\Omega$ to $1.75 \text{ k}\Omega$ with an average sensitivity of $\sim 1 \Omega/\text{Oe}$.

B. Magnetorelaxation Detection of Dried MNPs

To validate the concept, MNPs (Ocean Nano-Technologies SHS-30-01) with a core size of 30 nm were dried on the surface of the sensors. Part of the sensor chip was covered with epoxy to serve as a negative control. Fig. 11 shows two representative sensors, an active sensor with MNPs and a reference sensor without. The active sensor exhibited a relaxation signal with amplitude of $\sim 8 \text{ mV}$, while the reference sensor had no signal.

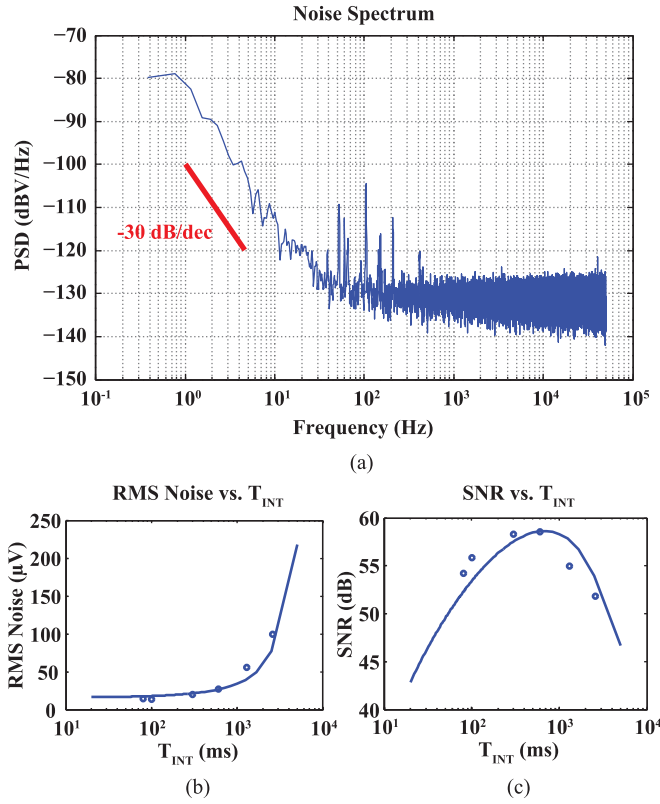


Fig. 12. Noise performance of system showing (a) Noise spectrum for $T_{\text{INT}} = 2.6$ s, (b) Simulated and measured RMS noise vs. T_{INT} , and (c) Simulated and measured SNR vs. T_{INT} .

C. Noise Analysis and Measurement

To optimize the SNR of the system, consider (3) and (4). R_D is required to be close to R_0 for current cancellation and therefore cannot be changed. Increasing C_F reduces both the signal and noise. Increasing V_B improves the SNR, but is limited by the breakdown voltage of the sensor. On the other hand, both C_F and V_B affect the integration time T_{INT} . While increasing T_{INT} increases the signal amplitude because there is more time to integrate the relaxation signal, it also increases the integrated noise by reducing the lower limit of integration f_{min} , where $f_{\text{min}} = 1/T_{\text{INT}}$. Hence, for a given sensor and MNP relaxation time, there exists an optimum T_{INT} .

The noise contributed by a sensor and op-amp were measured to be $i_{n,\text{Rs}} = 3.2 \text{ pA}/\sqrt{\text{Hz}}$ and $v_{n,\text{op}} = 2.7 \text{ nV}/\sqrt{\text{Hz}}$, respectively. Both noise sources have a $1/f$ corner frequency of ~ 1 kHz. Using (4) with these values, it turns out that $1/f$ noise is dominant below 10 kHz. Through subtracting integration curves in adjacent cycles, the noise in the integration phase can be extracted. To observe low frequency band, T_{INT} was extended to 2.6 s by decreasing V_B . In Fig. 12(a), although the high frequency band was limited by the ADC quantization noise, the low frequency band has a slope of -30 dB/dec , which matches with the theoretical analysis.

Due to the limitation on C_F , the relaxation signal under large T_{INT} but the same V_B cannot be measured. To analyze SNR for different T_{INT} to find the optimum, simulated relaxation

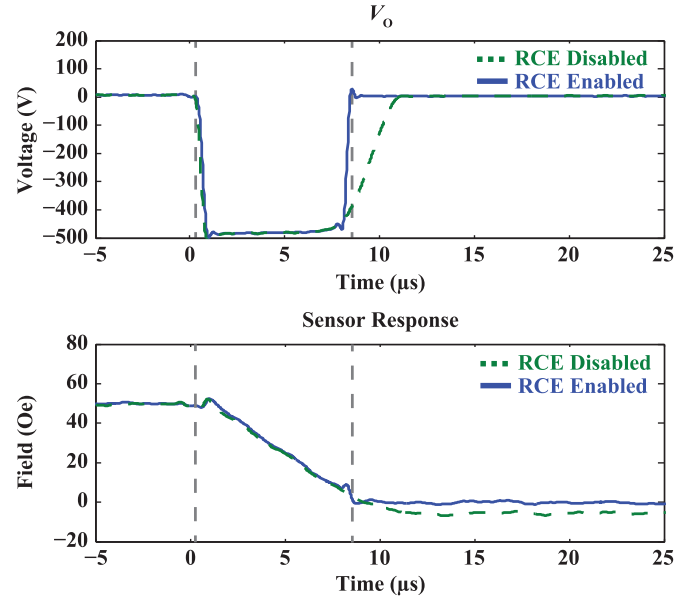


Fig. 13. Measurement results showing the reverse current elimination (RCE) technique.

signals were calculated based on (1). Since parameters C and t_C are unknown in (1), the simulated signal was first fitted to the measured signal for $T_{\text{INT}} = 100$ ms to extract the parameters. Then, the signal was estimated for different T_{INT} up to 5 s.

Since multiple cycles were used for signal averaging and T_{INT} would affect the number of cycles to average, a fixed measurement time of 60 s was applied for noise and SNR tests. It should be noted that MCDS requires two measurement cycles (one with field and one without field) to extract one processed cycle, and there is a gap of 0.6 s between integration curves in adjacent measurement cycles. Therefore, the sampled data in adjacent processed cycles were separated by the time of $2T_{\text{CYC}} = 2 \times (T_{\text{INT}} + 0.6)$ s. This time was long enough to ignore the correlation of $1/f$ noise, therefore both white noise and $1/f$ noise could be reduced through signal averaging. The simulated (lines) and measured (dots) noise and SNR vs. T_{INT} are shown in Fig. 12(b) and (c), respectively. The optimum T_{INT} was found to be 600 ms. Since the data with MCDS enabled is uncorrelated, signal averaging can be used to reduce the noise variance.

D. Electromagnet Driver Switch-off Response

The performance of the electromagnet driver switch-off response was measured using both a GMR sensor and an oscilloscope (Tektronics TDS2014C). By biasing the sensor with a DC current, the transient response of sensor was captured to monitor the magnetic field. The bandwidth of the GMR sensor, bias circuit, and oscilloscope was measured to be much higher than the switch-off response of the magnetic field.

To illustrate the performance improvement of the RCE technique, measurements with the RCE enabled and disabled are presented in Fig. 13. It can be observed that the RCE forced V_O back to 0 V rapidly when enabled. Thus, the reverse current,

TABLE I
COMPARISON OF DIFFERENT ELECTROMAGNETS

| Parameter | Coil 1 | Coil 2 |
|---|---|--|
| Inductance L | 1.5 mH | 80 μ H |
| Current I_0 | 2 A | 8 A |
| Physical Size (W \times L \times H) | 10 \times 8 \times 10 cm ³ | 15 \times 59 \times 23 cm ³ |
| Time Constant τ | 600 μ s | 1,600 μ s |
| Switch-off Time | 8.5 μ s | 1.4 μ s |
| Switch-off Speed | 5.9 Oe/ μ s | 35.7 Oe/ μ s |
| Improvement | 71 \times | 1,142 \times |

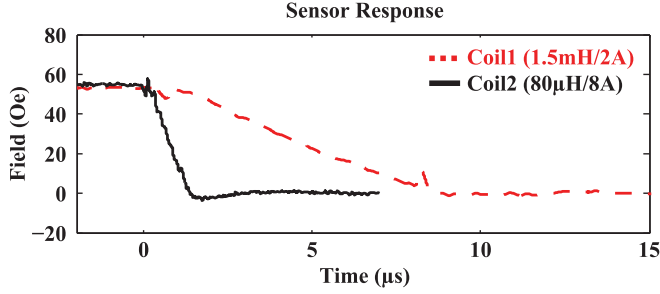


Fig. 14. Measurement results for comparison between two coils.

which caused a reverse magnetic field of up to -10 Oe and took 500μ s to dissipate, was fully removed. To verify the electromagnet optimization, two Helmholtz coils were custom designed, as summarized in Table I. The currents in both were set to achieve a magnetic field of 50 Oe. Fig. 14 shows the measurement results obtained from each of the electromagnets. The switch-off speed of Coil 2 (35.7 Oe/ μ s), which had lower inductance and higher current, was faster than Coil 1 (5.9 Oe/ μ s), supporting the electromagnet design principle stated earlier. However, Coil 2 requires $4\times$ higher current and a cooling fan for proper heat dissipation resulting in a larger physical size. Compared with the natural exponential decay time constant τ , the switch-off speeds for both coils were improved significantly enabling MNPs with relaxation times of only a few microseconds to be detected.

E. Magnetic Correlated Double Sampling Validation

The large voltage swing at V_{OUT} from V_{CM} to 0 V during the integration phase causes non-linearity from both the op-amp and the feedback capacitor. To illustrate this, a measurement was taken using a GMR sensor with no MNPs, as shown in Fig. 15(a). The non-linearity was extracted by subtracting the measured data from a linear fit of the curve. Without MCDS, the non-linearity is ~ 10 mV, which is comparable to the relaxation signal amplitude and similar in shape. After applying MCDS, the non-linearity is eliminated below the noise level. Similarly, the standard deviation of the sensor resistance on the same chip was measured to be 5.9Ω and reduced to 0.86 m Ω (99.98%) with MCDS.

While the non-linearity is constant over time, environmental variation such as temperature drift is not. This is illustrated in Fig. 15(b) where the temperature and amplitude of 7 sensors are plotted over time. 20μ L deionized (DI) water stored in the

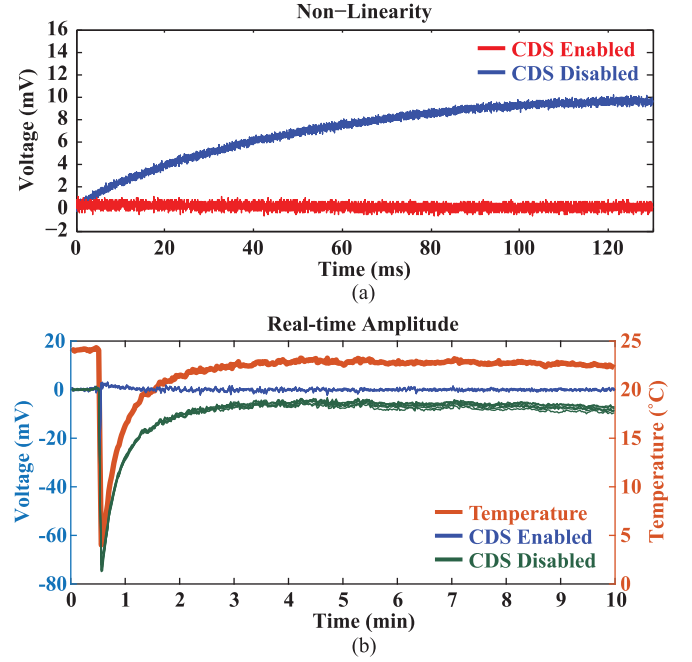


Fig. 15. (a) Non-linearity and (b) temperature sensitivity improvement from MCDS technique.

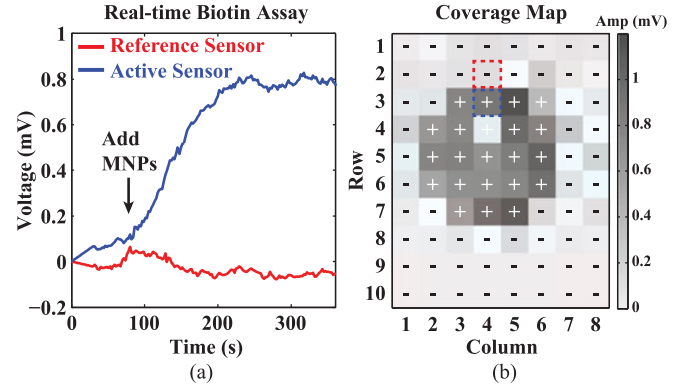


Fig. 16. (a) Real-time biotin assay experiment. (b) Post-assay full chip scan and signal amplitudes.

refrigerator (4°C) was added at $t = 40$ s, and the temperature was indirectly calculated from R_0 . The signals without MCDS show a strong correlation with the temperature whereas MCDS significantly reduces the temperature dependence. It should be noted that there is a single-point spike when the water was added since it occurred in the middle of an MCDS cycle.

F. Magnetic Immunoassay

Lastly, the experiment was extended from dried MNPs to an immunoassay. The sensor chip was partially functionalized with biotin and subsequently blocked with bovine serum albumin (BSA). The biotin on the active sensors binds with the streptavidin conjugated MNPs whereas the reference sensors were used to monitor non-specific binding. This experiment was performed with real-time measurement to observe the binding kinetics, as shown in Fig. 16(a). The MNPs were added at $t = 80$ s and

the experiment lasted until the binding plateaued. The sensors were then washed with phosphate-buffered saline (PBS) three times to remove unbound MNPs. The reference sensors and the washing step showed no change in signal amplitude indicating minimal non-specific binding. The full chip results are shown in Fig. 16(b) with the active sensors denoted by a + and the reference sensors by a -. The red and blue dotted squares indicate the reference sensor and the active sensor in the real-time measurement, respectively. The variation in amplitude for the active sensors was the result of the MNP surface coverage, which could be improved using individual spot functionalization versus the blanket functionalization procedure used here.

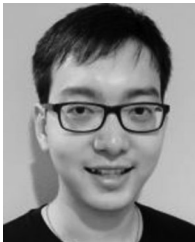
V. CONCLUSION

In this paper, a MRX biosensor system was demonstrated to detect the relaxation of 30 nm MNPs. A switched integrator was designed to integrate the relaxation signal with time, relaxing the bandwidth requirement of the ADC. An ultrafast electromagnet driver that actively dissipates the stored energy in the Helmholtz coil was designed to reduce the deadzone time. A magnetic correlated double sampling technique effectively eliminates the sensor-to-sensor variation, temperature drift, circuit offset and non-linearity. Measurement results for the electromagnet driver with a custom designed electromagnet show a fast switch-off response of $>5 \text{ Oe}/\mu\text{s}$. The experimental results of dried MNPs show that the relaxation of particles can be successfully detected and the biotin-BSA immunoassay demonstrates it is also possible to monitor the binding kinetics in real-time. This work lays the foundation for a fast and sensitive biosensing for point-of-care diagnostics platform.

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